Uganda Journal of Agricultural Sciences, 2001, 6: 43-46 Printed in Uganda. All rights reserved

### SHORT COMMUNICATION

### Gray leaf spot disease in Uganda

G. Bigirwa, K.F. Cardwell<sup>1</sup>, T. Sengooba, D.T. Kyetere, A. Nakayima and S.B. Kaboyo. Namulonge Agricultural and Animal Production Research Institute, P.O. Box 7084, Kampala, Uganda. <sup>1</sup>Plant Health Management Division, International Institute of Tropical Agriculture, Sub-station, Benin, BP os-0932, Cotonou.

### Abstract

Two studies were carried out to assess the status of gray leaf spot (GLS) disease of maize in Uganda. Results from three surveys of 1994, 1995 and 1996 conducted in 21 districts showed that GLS was widespread with high incidence and severity of 94% and 3.8 respectively. In importance it was ranking high followed by maize streak virus and turcicum leaf blight. Separation of individual pathogens from overall leaf area infection using backward regression showed that GLS accounted for over 60% of the leaf area infection. Work on varietal evaluation showed significant differences in reaction with three varieties; SC 627, SC 625 and SC Expt 2 showing high levels of resistance while H 622, SC 621 and PAN 6193 severely got affected.

Keywords: Gray leaf spot, severity, incidence, infection and pathogens

### Introduction

Several foliar maize diseases have been recorded to affect maize in Uganda. Recently however, gray leaf spot (GLS) caused by Cercospora zea-maydis Tehon & E.Y. Daniels has overtaken all the maize diseases in severity and spread. In the USA, GLS disease is reported to have become of economic since early 1970s (Beckman and Payne, 1982). Its increase and severity in the states has been linked with conservation tillage hence quite often referred to as a no-till disease (Rupe et al., 1982; de Nezareno et al., 1992). Continuous crop production, and extended periods of high relative humidity and dew points do favor the disease development and spread (de Nazareno et al., 1993). In Africa the disease was first reported in South Africa in 1988 (Ward et al., 1997) with a severe epidemic occurring in 1991/92 season (Gevers and Lake, 1994). Since then several African states started reporting severe epidemics. In the case of Uganda the first severe attack was noticed on farmers' fields in Mubende district during the first season of 1994 and this raised several concerns as many more fields and reports about the strange disease kept being reported to the National Maize Research Program at Namulonge Research Institute by extension agents and farmers who carried along specimens. Zimbabwe, and Kenya had the first epidemic in 1995/96 (Pixley, 1996) while Malawi and Cameroon experienced it in 1997 (P. Ngwira and Z. Ngoko personal communication).

The disease forms lesions which are gray to tan, rectangular  $2-5 \times 0.3-0.6$  cm long x 0.3-0.6 cm, and runs parallel to the leaf veins. During severe conditions, lesions may

coalesce and blight the entire leaf. The disease thrives best during prolonged periods of hot and humid weather and is potentially more severe in fields where maize follows maize and where reduced tillage practices are used (de Nazareno et al., 1993). The fungus within the infested debris produces conidia, which are eventually blown by wind and thus infecting the new crop. Varying yield losses have been reported depending on the location and genotype. Ward et al., (1993) estimated a yield loss of 88% in South Africa while Saghi Maroof et al., (1996) estimated the loss in the USA to range between 10-50%. Gray leaf spot disease on the other hand is known to an environmentally dependant disease to the extent that even if the inoculum is present but once the weather conditions are not right the will be no disease development and this is the reason why in some seasons or years no disease is noticed (Beckman and Payne, 1982).

Following the unexplained epidemic in Uganda during the year 1994 and subsequent seasons it was found necessary to initiate some research activities to address the problem. The objective of the study therefore was to carry out a survey to ascertain the incidence, severity and distribution; and at the same time evaluate the elite materials and commercial varieties available.

### Materials and methods

A countrywide survey of maize diseases in Uganda was carried out during the second seasons of 1994 and 1995, and in the first season of 1996. The districts surveyed included Mukono, Jinja, Iganga, Tororo, Mbale, Paliisa, Soroti, Apac,

Lira, Masindi, Hoima, Nakasongola, Luwero, Mpigi, Mubende, Kiboga, Masaka, Mbarara, Bushenyi, Kabarole and Kasese. Field sampling was made along main road systems by stopping every 20 km. In each field 40 plants were selected, i.e.10 in each of the 4 different portions of the field from which incidence and severity were taken. A scale of 1-5 was used to determine severity, where 1= no or very few lesions and 5= many lesions and leaves severely blighted. Total leaf area was visually estimated and expressed in percentages. Nitrogen deficiency was also visually estimated by assessing the amount of foliage showing deficiency symptoms. A similar scale of 1-5 was used; 1= no symptoms and 5= severe symptoms. In 1996 an intensive survey was carried out in Iganga district covering 3 counties (Luuka, Kigulu and Bunya). The same protocol was repeated except stops were being made every 5 km.

In an attempt to identify resistant varieties, 14 local and foreign hybrids mostly from private seed companies and 2 open pollinated varieties were evaluated in 2 locations; Namulonge and Kamenyamiggo. Test materials were planted in 2 row 5 metre plots in a randomized complete block design with 4 replications at a plant population of 53,000 plants per hectare. At V6 stage (Ritchie et al., 1989), seedlings were inoculated with dry, ground infected leaves by placing a pinch into the whorls. Inoculation was repeated after one week. Eight plants were randomly selected and tagged from each plot for the assessment of percent ear leaf area affected (PLAA) as described by Freppon et al. (1996). Disease assessment commenced at R1 stage. A total of five assessments were done at an interval of 7-10 days. Severity was recorded at green maturity, using 1- scale as described in the survey.

Differences in severity and PLAA were determined by analysis of variance (ANOVA) and mean separation was based on Fisher's least significant differences procedure (LSD) at 5% level of probability. All analyses were performed with MSTATC statistical analysis software (Feed *et al.*, 1988).

### **Results and discussion**

Five diseases were commonly found in almost all locations surveyed and with varying degrees of severity and incidence; GLS, maize streak virus, northern leaf blight (Exserohilum turcicum), sternocarpella leaf spot (Sternocarpella macrospora) and southern leaf blight (Bipolaris maydis). Of all theses, GLS ranked high in severity, distribution and incidence in all the three seasons with an average incidence of 90.6% and severity of 3.4 (Table 1). It was followed by maize streak and northern leaf blight whose average incidence and severity was 14.9%, 1.8 and 9.7%, 1.4 respectively. Gray leaf spot incidence was highest in 1996 for instance in Bunya all 30 fields surveyed had GLS with an average severity of 3.9, in Luuka incidence from the 27 fields was 91.8% and a severity of 3.9 while in Kigulu incidence was 87.2% from 19 fields with an average severity of 3.3. Many plants were found lodging and others without cobs. Brief interaction with farmers showed that they had come to know that the bizarre appearance of their fields was due to some disease or pest although in 1994 and 1995 they thought it was due to drought. Sporulation of C. zeae-maydis was quite evident on many leaves. Effects of nitrogen deficiency were observed in many fields for instance, in 1994 its incidence was 9.2% while in 1996 it was recorded as 7.1%

Separation of individual pathogens from the overall leaf area infection using backward regression on data of 1996 showed that GLS was responsible for 60% of leaf infection observed (Table 2). This clearly confirms the fact that GLS was the most devastating disease then. It was followed by northern leaf blight and maize streak virus.

Disease	Percentage of fields infected			Disease severity (1-5 scale)		
	1994	1995	1996	1994	1995	1996
GLS.	87.6	89.3	94.8	3.2	3.8	3.3
MSV	17.6	12.9	14.1	1.8	1.6	1.9
NLB	12.3	7.0	9.9	1.5	1.3	1.5
Smac	3.7	4.7	5.6	1.0	1.2	1.5
Ndef	9.2	3.8	7.1	1.2	1.1	1.2
SLB	1.4	1.6	2.3	1.0	1.0	1.1
Mean	22.0	19.9	22.3	1.6	1.7	1.8
LSD (0.05)	15.3	11.6	14.4	0.3	0.4	0.4

Table 1. Foliar disease severity and percentage of fields infected in Uganda between 1994-1996.

<sup>a</sup>GLS=gray leaf spot, MSV= maize streak virus, NLB= northern leaf blight, Smac= sternocarpella macrospora, Ndef= nitrogen deficiency and SLB= southern leaf blight.

Varieties evaluated against *C. zeae-maydis* in the two locations showed varying levels of susceptibility with clear significant (P<0.01) differences. Pattern of reaction was consistent to the extent that varieties found either susceptible in Namulonge were equally susceptible in Masaka. The same

### Table 2. Contribution of individual pathogens to overall leaf area infection (y) in Iganga, during the second season of 1996, using backward regression.

Variable	b	т	Sign. T
Smac <sup>a</sup>	4.77	2.26	0.000
NLB	4.31	7.69	0.007
MSV	9.24	4.53	0.000
GLS	15.26	19.82	0.000
Ndef	5.83	3.47	0.004
Intercept R2 = 0.68	-23.75	-6.18	0.000

\*Smac= sternocarpella macrospora, NLB= northern leaf blight, MSV= maize streak virus, GLS= gray leaf spot and Ndef= nitrogen deficiency.

Table 3. Severity and percent ear leaf area affected (PLAA) by gray leaf area spot disease on maize genotypes at Kamenyamiggo and Namulonge during the second season of 1997

Maize	Kamenya	imigo	Namulonge	
genotype	Severityª (1-5 scale)	PLAA <sup>B</sup>	Severity (1-5 scale)	PLAA
NZ 4	4.0	45.0	3.6	44.4
Longe 1	3.9	40.5	3.0	40.8
H 622	4.7	41.3	4.5	45.0
Katumani	3.9	42.0	3.2	43.9
H 512	3.7	40.3	3.6	45.0
H 511	3.0	43.0	3.1	44.7
PAN 6193	4.3	42.5	4.4	45.0
LP 16	3.1	42.3	3.3	43.9
PAN 67	3.5	40.5	3.4	44.4
SC 621	3.1	36.6	3.1	38.1
SC 625	1.2	10.1	1.2	7.9
SC 627	1.6	14.9	1.8	17.9
SC Expt2	1.0	7.9	1.2	9.7
NZ 1	4.7	45.0	4.7	45.0
NZ 2	3.9	42.5	3.5	40.7
NZ 3	3.5	41.9	2.9	39.9
EVS 885°		- ur,	3.2	43.6
Mean	3.3	36.5	3.2	37.8
LSD (0.05)	0.6	10.1	3.2	18.6

<sup>a</sup>Severity taken 90 days after inoculation <sup>b</sup>Percenta ear leaf area taken 90 days after i..oculation<sup>c</sup>Genotype EVS 855 was not tested at Kamenyamiggo. is true for the resistant varieties. Three hybrids SC 625, SC 627 and SC Expt 2 were exceptionally resistant as reflected by their severity scores and percent leaf area affected (Table 3) while PAN 6193, SC 621, NZ 1 and H 622 were extremely susceptible. The present commercial variety Longe I was noted to be susceptible a similar situation found in the field. These findings were gratifying because there was hope that some varieties were resistant not only to GLS but to other common diseases in the country particularly maize streak virus and northern leaf blight; and in addition they were high yielders. For Longe I the commercial variety research focus was going to aim at improving its level of resistance using the recurrent selection method.

The study clearly showed that GLS was widely distributed in the country and with a potential to cause considerable yield losses. As to whether it would remain a big problem, this would be determined by a number of factors like failure to avail resistant varieties and put in place feasible integrated control management options. In addition, since the disease is known to be weather dependant (Beckman and Payne, 1982; Rupe *et al.*, 1982 and Freppon *et al.*, 1998), the common weather patterns frequently experienced are bound to play a significant role on its impact.

### References

- Beckman, P.M. and Payne, G.A., 1982. External growth, penetration and development of *Cercospora zeae-maydis* in corn leaves. *Phytopathology*, 72:810-815.
- de Nazareno, N.R.X., Lipps, P.E. and Madden, L.V. 1992. Survival of Cercosporo zeae-maydis in corn residue in Ohio. Plant Disease, 76:560-563.
- de Nazareno, N.R.X., Lipps, P.E. and Madden, L.V., 1993. Effect of levels of corn residue on epidemiology of gray leaf spot in Ohio. *Plant Disease*, 77: 67-70
- Freed, R., Eisensmith, S.P., Goetz, S., Reicosky, D., Smail, V.W. and Wolberg, P., 1988. Mstatc. A microcomputer program for the design, management, and analysis of agricultural research experiments. Michigan State University pp 1-29.
- Freppon, J.T., Pratt, R.C. and Lipps, P.E., 1996. Chlorotic lesion response of maize to *Cercospora zeae-maydis* and its effect on gray leaf spot disease. *Phytopathology*, 86:733-738.
- Gevers, H.O. and Late, J.K. 1994. Diallel cross analysis of resistance to gray leaf spot in maize. Plant Dis. 78:379-383.
- Ritchie, S.W., Hanway, J.J. and Benson, G.O., 1989. How a corn plant develops. Iowa State University Special Report 48. Pages 24
- Pixley, K.V., 1996. CIMMYT mid-altitude breeding program report of activities during 1995/97. Chapter 2. In: D.C. Jewell, K.V. Pixley, M. Banziger, S.R. Waddington, T.S. Payne and B.T. Zambezi (eds) *Annual Report, CIMMYT-*Zimbabwe, Harare, Zimbabwe pp 6-32.
- Rupe, J.C., Siegel, M.R. and Hartman, J.R., 1982. Influence of environment and plant maturity on gray leaf spot of corn caused by *Cercospora zeae-maydis*. *Phytopathology*, 72:1587-1591.

Saghai Maroof, M.A., Yu, Y.G., Xiang, Z.X., Sromberg, E.L. and Rufener, G.K., 1996. Identification of quantitative trait loci controlling resistance to gray leaf spot disease in maize. *Theor. Appl. Genet.*, 93:539-546 Ward, J.M.J., Laing, M.D. and Cairns, A.L.P., 1997. Management practices to reduce gray leaf spot of maize. *Crop Sci.*, 37: 1257-1262.

# **Guide for Authors**

**1.Types of contribution.** This could be: (i) original full papers (regular papers), (ii) invited review articles, (iii) short communications, and (iv) views and ideas. Original papers should report the results of original research. The material should not have been previously published elsewhere, except in a preliminary form. Reviews should cover a part of the subject of active current interest.

A short communication is a concise, but complete, description of a limited investigation, which will not be included in a later paper. Short Communications should be as completely documented, both by reference to the literature and description of the experimental procedures employed, as a regular paper.

The section views and ideas offers comment or useful critique on material published in the journal or on relevant issues. Contributions to this section should not occupy more than 2 printed pages.

### 2.Submission of manuscripts

The article should be original and not being considered, for publication elsewhere. Hard-copies are sufficient for the initial submission of manuscripts. However, for the processing of accepted papers, electronic versions(diskettes) are preferred. After final acceptance, your diskette plus two, final and exactly matching printed versions should be submitted together. Label the disk with the name of the computer and wordprocessor package used, your name, and the name of the file on the diskette.

### 4. Preparation of manuscripts

a) Manuscripts should be written in English;

 b) Submit the original and two copies of your manuscript.
 Enclose the original illustrations and two sets of photocopies (three prints of any photographs);

c) Manuscripts should be typed on one side of the paper with double spacing throughout. Every page of the manuscript, including the title page, references, tables, etc. should be numbered;

d) Manuscripts in general should be organized in the following order: Title (should be clear, descriptive and not too long); Name(s) of author(s); Complete postal address(es) of affiliations; Abstract; Key words (indexing terms); Introduction, Material studied, area descriptions, methods, techniques; Results; Discussion; Conclusion; Acknowledgements and any additional information concerning research grants, etc; References; Tables; Figures; Legends.

e) In typing the manuscript, titles and subtitles should not be run within the text. They should be typed on a separate line, without indentation. Use lower-case lettertype.
f) SI units should be used.

5. Abstract. The abstract should be clear, descriptive and not longer than 200 words.

6. Tables. Place each table on a separate page and lebel them clearly with a number and title; double-space all material; omit vertical lines. Units of measure should be metric, with two places of decimals. Base-dates for index numbers, geographical area covered and sources be clearly stated. Statistical differences, letters for the 5% (<0.05) and 1% (<0.01) significance levels should be indicated. Avoid footnotes. Authors are fully responsible for the accuracy of the data and for checking their proofs, but whenever they feel that the referee would have difficulty

in testing the derivation of their statistics, they should provide supplementary notes on the method used, which will not be published.

7. Figures and illustrations. Figures should be clearly drawn, with clearly marked axes and must be numbered. Lettering on the figures must be sharp and dark enough to withstand reduction. Figures must be cited in the text, submitted on separate sheets and accompanied by the basic statistics required for their preparation, when appropriate. In short, send the illustration (say bar graph) with the tables from which the graph was derived. The editors may prefer your graph presented as a pie chart or as a table. Illustrations should be in black and white, if colour is (absolutely) necessary, indicate in the covering letter.

#### 8.References

a)All publications cited in the text should be presented in a list of references following the text of the manuscript. Carefully check the manuscript to ensure that the spelling of author's names and dates are exactly the same in the text as in the reference list.

b) In the text refer to the author's name (without initial) and year of publication. Examples: Mukiibi: "(1982) has shown that ...." "This is in agreement with results obtained later (Bekunda, 1990)".

c) If reference is made in the text to a publication written by more than two authors the name of the first author should be used followed by "et al.". This indication, however, should never be used in the list of references. In this list, mention the names of first author and co-authors. d) Chronologically arrange references cited together in the text. Arrange the list of references alphabetically on authors' names, and chronologically per author. If an author's name in the list is also mentioned with co-authors the following order should be used: publications of the single author, arranged according to publication dates publications of the same author with one co-author publications of the same author(s) in the same year should be listed as 1974a, 1974b, etc.

e) Arrange your references as shown:

(i) For books, write as:

Gaugh, Jr., H.G., 1992. Statistical Analysis of Regional Yield Trials. Elsevier, Amsterdam, 278 pp.

Stover, R.H. and Simmonds, N.W., 1987. *Bananas.* Tropical Agriculture Series. Longman, Singapore, 468 pp. (ii) For periodicals.

Bekunda, M.A., Smethurst, P.J., Khanna, P.K. and Willet, J.R., 1990. Effects of post-harvest residue management on labile soil phosphorus in a *Pinus radiator* Plantation, *Forest Ecology and Management*, 38:13-25.

(iii) For edited symposia, special issues, books, etc. published in a periodical, write as:

Swift, M.J., Bohren, L., Izac, A.M. and Woomer, P.L., 1994 Biological management of tropical soil: Intergrating process research and farm practice. In: P.L. Woomer and M.J. Swift (Editors), *The Biological Management of Tropical Soil Fertility*. John Wiley, Chickester, UK, pp. 209-227.

f) Titles of periodicals in the list of references should be in full.

g) Work accepted for publication but not yet published should be referred to as "in press".

h) References concerning unpublished data and "personal communications" should not be cited in the reference list but may be mentioned in the text.

# **Guide for Authors Continued**

### 9. Formulae

a) Leave ample space around the formulae.

b) Subscripts and superscripts should be clear.

c) Clearly show the difference between letters and figures that resemble, e.g. zero (0) and letter O, and one (1) and the letter I.

d) Give the meaning of all symbols immediately after the equation in which they are first used.

e) Equations should be numbered serially at the righthand side in parentheses. In general only equations explicitly denoted by exp.

f) The use of fractional powers instead of root signs is recommended. Also powers of e are often more conveniently denoted by exp.

g) Levels of statistical significance which can be mentioned without further explanation are \*p<0.05, \*\*p<0.001 and \*\*\*p<0.001.

h) In chemical formulae, valence of ions should be given as e.g.  $Mg^{2+}$  and  $SO_4^{-2-}$ , not as  $Mg^{--}$  or  $SO_4^{--}$ .

i) Isotope numbers should precede the symbols, e.g. <sup>14</sup>C. j) The repeated writing of chemical formulea in the text is to be avoided where reasonably possible; instead, the name of the compound should be given in full. Exceptions

and the second second

may be made in the case of a very long name occurring very frequently or in the case of a compound being described as the end product of a gravimetric determination (e.g. phosphate as  $P_2 O_s$ .

k) Use e.g. kg ha' instead of kg/ha, etc.

### 10. Nomenclature

a) With the exception of common domestic animals, identify all biotica (crops, plants, insects, birds, mammals, etc.) by their scientific names when the English term is first used.

b) All biocides and other organic compounds must be identified by their Geneva names when first used in the text. Active ingredients of all formulations should be likewise identified.

c) For chemical nomenclature, the conventions of the International Union of Pure and Applied Chemistry and the official recommendations of the IUPAC-IUB Combined Commission on Biochemical Nomenclature should be followed.

### 11. Offprints

Could be ordered in an off print order form.

## **Order Form**

### Uganda Journal of Agricultural Sciences (UJAS)

### Subscription Information for 1 Year

	Local	Africa	Elsewhere
Institutions	\$ 40	\$ 80	\$ 120
Individuals	\$ 20	\$ 40	\$ 60

If you wish to be a subscriber please tick in the approprate box and return this slip with the information filled below:

Full name: Prof./Dr/Mr/Ms
Designation
Organisation/Institution
Address
CountryPost/Zip code
E-mail
Tel:
Fax:

Signature.....Date.....

NB: Prices in Uganda Shillings are determined by the Dollar exchange rate at that particular time

All payments should be by Bank draft cheques payabale to "Uganda Journal of Agricultural Sciences(UJAS)", National Agricultural Research Organisation(NARO), P.O. Box 295, Entebbe, Uganda or sent by telegraphic transfer to "UJAS, Acc. No.0119431-001, Nile Bank Ltd, Main Branch, Plot No 22 Jinja Road, Kampala, Uganda.

# **Uganda Journal of Agricultural Sciences**

A Multi-Disciplinary Journal Devoted to the Study and Promotion of Agriculture and Natural Resources, Livestock and Fisheries Development

# (Volume 6 Issue 1)

# CONTENTS

Assessing tropical moist forest conditions: the case of Mengo forests. Gombya-Ssembajjwe, William, S.	1-5
On-farm tree planting and tree diversity in the Kigezi Highlands and Mabira Buffer Zones. Joseph Obua, Geoffrey Muhanguzi and Thomas Raussen.	7-12
The response of broiler chicks to dietary serena sorghum (Sorghum bicolor). M.W. Okot and S.N. Mujabi.	13-18
A case report of <i>Histomonas</i> infection in chicken from Pallisa District in Uganda. J. Illango, G. Musisi, A. Etoori.	19-20
A review of climbing bean variety evaluation and adoption in south western Uganda. P. Tukamuhabwa, H. Gridley, B. Kayiwa and C. Niringiye.	21-25
A Century of banana research and development in Uganda 1898-1998. W.K. Tushemereirwe.	27-36
Inventory of agricultural biotechnology research capacity in Uganda. Thomas Braunschweig and Theresa Ssengooba.	37-41
Gray leaf spot disease in Uganda. G. Bigirwa, K.F. Cardwell, T. Sengooba, D.T. Kyetere, A. Nakayima and S.B. Kaboyo.	43-46